

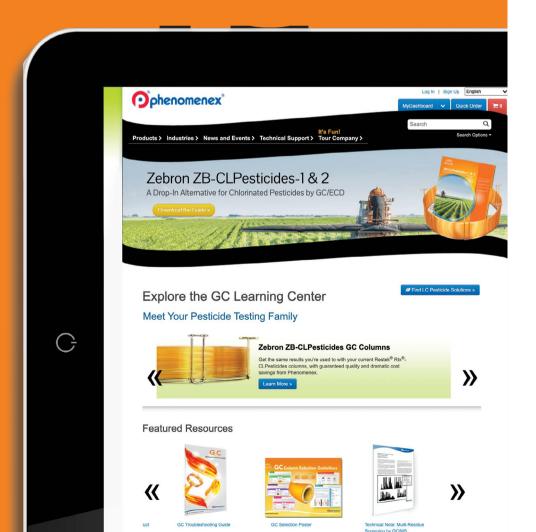


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GC Troubleshooting Guide

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Column Conditioning

System Conditioning

BEFORE YOU START

Goals & Approaches

An important first step in troubleshooting is understanding the problem. This is best approached systematically; once you have a good understanding of what is causing the problem, it will be easier to implement a logical solution. Understanding the problem can also allow you to alter analysis or maintenance habits to avoid the problem in the future. Prevention is usually the most cost effective solution!

Common Troubleshooting Approaches



Changing everything relating to the problem may allow a remedy, but won't allow you to fully understand the solution. In the long run, this may be more costly than other approaches. Make An Educated Guess

Narrowing down or eliminating parts of the process may give a good starting point to determine the cause of the problem, but does not necessarily allow you to solve or prevent the problem in the future.



A systematic approach includes checking the chromatographic system from one end to the other, isolating the problem, learning what went wrong, solving the problem, and then preventing it. This is the recommended approach!

Problem Prevention

Many GC problems can be prevented if the column and system are maintained routinely. *Problem Prevention* (see p. 32) outlines maintenance practices that will reduce the frequency of common issues. These suggestions should be modified to fit your GC column and instrument, and then made a regular part of your laboratory routine.

Troubleshooting Tools

What To Have On Hand

Have your instrument manual and these diagnostic tools at hand:

- Flow meter with a range of 10 to 500 mL/minute
- New syringes
- Non-retained, detectable compound such as methane or propane
- Septa, ferrules, inlet liners, and other consumables
- Electronic leak detector
- Reference sample
- · Reference column with known performance

BEFORE YOU START

Tips For Effective Troubleshooting

- When troubleshooting, remember to look at recent changes in the system, especially if the system was working
 previously. Was there something that changed and may be causing the problems now? Can you undo the
 changes and go back to the original performance?
- Try to isolate the problem to one specific cause to minimize the changes that have to be made to the system
 that may result in other malfunctions. This will also make it easier to prevent the problem in the future and
 shorten troubleshooting times if a similar problem does occur in the future.
- Remember to check every part of the process. Don't overlook the obvious. If you are not getting peaks
 for example, the makeup flow to the detector may be off or the syringe may be clogged. Has the sample
 preparation method been altered? Verify your samples on another instrument if possible.
- Keep good records of the troubleshooting process closely observe and note operating parameters
 (temperatures, flow rates, columns used, etc.). Reliable system maintenance records (inlet liner changes,
 detector cleanings, etc.) are also important.

For Additional Help

- The operator's and service manuals for the instrument should be consulted. These contain exploded diagrams, troubleshooting procedures for specific models, and part numbers to help you order replacement parts.
- Other people in the lab may have had experience solving a problem that is giving you trouble; they can be a helpful resource.
- The manufacturer of your instrument can help you. Most GC manufacturers offer free technical support to their
 customers. Phenomenex has experienced technical consultants who can assist you with almost any problem. We
 welcome your phone calls or emails!

BASELINE PROBLEMS

BASELINE PROBLEMS

Bleed

Symptom



Possible Cause

Improper column conditioning.

Contaminated column.

Contaminated injector.

Leak in system causing column oxidation.

Septum is not conditioned.

Septum core is present in the flow path.

Suggested Remedy

Properly condition the column.

There are several options:

- Trim the column
- Bake out the column
- Solvent rinse the column
- Replace the column

Perform inlet maintenance – clean the injector, replace the inlet liner, replace glass wool. See inlet maintenance.

Check for leaks in the system. Tighten or replace connections; replace seals and filters. If column is severely damaged, replace.

See column installation.

Condition septum prior to analysis or use pre-conditioned septum. Check septa temperature rating – should be sufficient to run at method temperatures.

Remove septum core from the inlet. Check septum nut and make sure it is not over tightened. Inspect injector syringe for bent or blunt tip and replace as necessary.

Drift Slow movement of the baseline in one direction (either up or down).

Symptom



Upward

Possible Cause

Downward drift for a few minutes is normal after installing a new column.

Unequilibrated detector or oven.

Downward drift is frequently due to the "bakeout" of contaminants from the detector or other parts of the GC.

Excessive damage to the stationary phase of the GC column.

Drift in gas flow rates.

Suggested Remedy

Increase the oven temperature to the maximum continuous operating temperature for the column. Maintain that temperature until a flat baseline is observed. If the detector signal continues to raise or does not drop in 10 minutes, immediately cool the column and check for leaks.

Allow sufficient time for (temperature) equilibration of the detector or oven.

Clean out contamination. See detector maintenance.

Determine the cause of the damage. It may be due to impurities in the carrier gas or to excessive temperatures. Replace column.

See column installation.

Clean or replace flow or pressure regulator(s). Adjust pressure.

See column installation.

BASELINE PROBLEMS

Noise Rapid, random movement of the signal amplitude.

Symptom



Possible Cause

The column may be inserted too far into the flame of an FID, NPD or FPD detector.

An air leak can result in noise in ECD and TCD detectors.

Incorrect combustion gases or flow rates can generate noise in FID, NPD, or FPD detectors.

Contaminated injector.

Contaminated column.

Drift in gas flow rates.

Defective detector board.

Suggested Remedy

Reinstall the column. Be sure to insert the column into the detector exactly the correct distance specified in the instrument manual.

See column installation.

Eliminate the leak.

Be sure your gases are the proper grade, as well as clean and dry. Reset the flow rates of the gases to their proper values.

Clean injector. Replace inlet liner. Replace glass wool. See injector maintenance

Bake out the column.Cut off first 4 inches of column. Solvent rinse or replace column.

Clean and/or replace parts as necessary.

See detector maintenance.

Consult GC instrument manufacturer.

BASELINE PROBLEMS

Offset Sudden unexplained changes in baseline position.

Symptom



Possible Cause

Line voltage changes.

Poor electrical connections.

Contaminated injector.

Contaminated column.

Column inserted too far into the flame of FID, NPD, or FPD detectors.

Contaminated detector.

Gas generator cycle.

Suggested Remedy

Monitor line voltage for correlation with offset. If correlation is found, install voltage regulator.

Check electrical connections. Tighten any loose connections. Clean any dirty or corroded connections.

Clean injector. Replace inlet liner. Replace glass wool. See injector maintenance.

Bake out the column. Cut off the first 4 inches of column. Solvent rinse or replace the column. See column installation

Reinstall the column. Be sure to insert the column into the detector exactly the correct distance specified in the instrument manual.

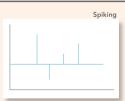
See column installation.

Clean the detector. See detector maintenance.

Baseline fluctuations can occur as the generator turns on and off. Add a tank with the appropriate volume after the generator to buffer any pressure changes.

Spiking Peaks with no width, either positive or negative.

Symptom



Possible Cause

Electrical disturbances entering the chromatogram through power cables, even shielded cables.

Particulate matter passing through the detector.

Pressure may build up and gas may escape through a seal and thus reduce the pressure below the point where the escape occurs. If this is the cause, the frequency of spikes will be pressure-dependent.

Loose, dirty, or corroded electrical connections in the detector or at connections along the signal path can cause spiking.

Suggested Remedy

Try to correlate spikes with events in equipment near the chromatogram. Periodicity is often a clue. Turn off equipment or move it. If necessary, install a voltage regulator.

Clean the detector and eliminate the source of particles. A clean hydrogen flame is invisible. Most organic matter generates a yellow flame. See detector maintenance.

Fix leaking seal.

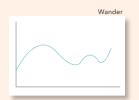
Check electrical connections. Tighten any loose connections. Clean any dirty or corroded connections. Replace badly corroded FID parts.



BASELINE PROBLEMS

Wander Low frequency noise.

Symptom



Possible Cause

Baseline wandering may be caused by changes in environmental conditions such as temperature or line voltage.

Inadequate temperature control. Check if variations can be correlated with changes in the baseline position.

Wandering while using isothermal conditions may be due to contaminated carrier gas.

Contaminated injector.

Contaminated column.

Poor control of gas flow rates.

Suggested Remedy

Try to correlate the wandering with environmental parameters. If a correlation is observed, you will know what to do. Good luck.

Measure detector temperature. Check detector, if TCD is used.

Change the carrier gas or the gas purification traps.

Clean injector. Replace inlet liner. Replace glass wool. See injecor maintenance.

Bake out the column. Cut off first 4 inches of column. Solvent rinse or replace column.

Clean or change flow controller(s).



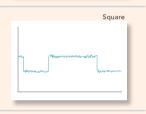
BASELINE PROBLEMS

Waves Baseline oscillations different from typical noise.

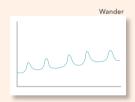
Symptom











Possible Cause

Detector related problem.

Gas pressure fluctuations.

Unbalanced column-switching or gas-sampling valves (for TCD

AC power fluctuations; interference from other equipment

Excessive column bleed.

Oxygen contamination is degrading the column phase.

Contamination during isothermal parts of a run.

Suggested Remedy

Baking the detector at the maximum temperature (450°C for FID) for 30 min to 1 hour may provide temporary relief. For a lasting solution, physically clean the detector.

Storage tank pressure varies, causing dips in flow. Adding a dual stage regulator can minimize pressure fluctuations and help to alleviate the problem.

Measure, check, and set flows accurately.

Use a dedicated AC source with sufficient power.

Check max column temperature, re-condition column. If column is damaged beyond repair, replace

Install or check oxygen traps. Check system for oxygen leaks.

The column is separating the contaminants (commonly siloxanes or hydrocarbons) as peaks. Changing samples, wash solvents, liner, gold seal, and sometimes the syringe may be required to eliminate the contamination.

PEAK SHAPE PROBLEMS

Reduced Size Some or all peaks are reduced in size.

Symptom



Possible Cause

Defective or plugged syringe.

"Blown" septum or other massive leaks at the inlet or with carrier gas flow. Poor peak shapes usually result from bad leaks.

Purge flow or split ratio too

Injector and/or column temperature too low for high molecular weight or low volatility samples.

NPD detector may be coated with silicon dioxide due to column bleed or residual derivatization reagents.

NPD damage by loss of rubidium salt as a result of exposure to overheating, heating in the absence of clean gas flow, or humidity.

For splitless injection, if the split vent is closed for too short a period of time or if the initial column temperature is too high, this may hinder refocusing of the sample.

Detector-sample mismatch.

Inadequate signal amplification.

Sample invalidity.

Activity in the inlet liner or column if the reduced peak is an active compound.

Leak in the injector if the reduced peak is a more volatile

Initial temperature too high for splitless or on-column injections.

Analytes are decomposing or breaking down for active or thermally labile compounds.

Suggested Remedy

Try a new or proven syringe.

Find and fix leaks and adjust gas flow. See column installation.

Adjust gas flow rates.

Increase injector and/or column temperature(s).

Replace the active element. Avoid exposure to silicon containing compounds.

Replace the active element. Turn off detector whenever the gas flow is interrupted. Avoid overheating. Keep element warm (150°C) when not in use. Use a desiccator for extended storage.

Increase the time the split vent is closed. Decrease the initial column temperature or use a less volatile solvent so that the initial temperature is below the boiling point of the solvent.

Ensure that the detector will respond to the analytes.

Check output signal levels.

Check sample concentration and stability.

Clean or replace the inlet liner. Ensure an inert column is used. If necessary, replace the column.

Find or repair the leaks and adjust gas flow.

Lower the initial column temperature.

Use a higher boiling solvent.

Check the integrity of the sample.

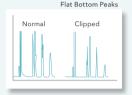
For thermal lability, lower the temperature and use on-column injection, a column with thinner stationary phase, a shorter column lengths, or a higher carrier gas flow rate. For active compounds, ensure an inert column is used. If necessary, replace the column.

Some Peaks Reduced

Clipped/Flat Peaks are clipped and flat at either the top or the bottom.

Symptom





Possible Cause

Detector overload. The broad peaks may have a rounded top or even valleys in the top.

Overload of the signal processing electronics. The peaks are clipped with flat tops.

Detector, recorder, or integrator set too low; detector drifted below zero.

Suggested Remedy

Reduce sample volume, dilute with solvent, or increase or add a split flow.

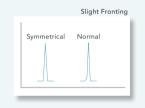
Attenuate detector output reduce sample volume, or add a split flow

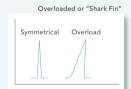
Correctly set zero. Reconnect leads from recorder and set zero of recorder baseline to ~5% of full scale. Check the integrator threshold and adjust accordingly.

Use an auto-zero function.

Fronting Moderate to severe asymmetry towards the front or left side of the peak.

Symptom





Possible Cause

Improper column installation.

Sample is condensing in the injector or column.

Column is overloaded as a result of injection volume and split

Polarity mismatch.

Suggested Remedy

Reinstall the column. See column installation

Check injector and oven temperature with an accurate thermometer. If accurate, increase temperature as necessary, but do not exceed the maximum temperature limit of the column.

Reduce the injection volume; add or increase split flow.

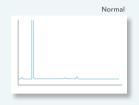
Use a column with greater capacity. Columns with larger diameter or thicker stationary phase coatings generally have larger sample capacities; however, resolution may be reduced.

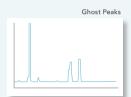
Polar compounds will have lower concentration capacity on a non-polar phase and vice-versa. Choose a phase with the appropriate polarity and selectivity for your sample.

PEAK SHAPE PROBLEMS

Ghost Peaks Peaks observed when no sample has been introduced into the system.

Symptom





Possible Cause

Remnants of previous samples in the inlet or column are most likely to occur when increasing inlet or column temperature(s).

Sample expanded to exceed the volume of the injector liner. These vapors may come in contact with colder spots, such as the septum and gas inlets to the injector. Less volatile components may condense. These condensates may vaporize later and interfere with subsequent analyses, sometimes producing "ghost peaks".

Bleed from the septum or fragments of the septum lodged in the inlet or liner

Syringe contamination.

Suggested Remedy

Increase the final temperature and lengthen the run time to allow for the complete elution of previous samples. If ghost peaks continue to occur, clean the inlet.

See injector maintenance.

Condition the column at a higher temperature that is still lower than the maximum isothermal limit for the column. Cut 4 inches off the inlet end of the column and/or reverse it (endfor-end) before reconditioning. Solvent rinse or replace the column.

See column installation.

Minimize backflash by using:

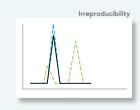
- a septum purge
- small injection volumes
- large inlet liners
- optimal injector temperatures
- pulsed pressure programming
- increased split flow

Clean the inlet. Replace the inlet liner or glass wool, and septum.

Replace the syringe.

Irreproducibility Peak heights, areas, or retention times are inconsistent from injection to injection

Symptom



Possible Cause

Inconsistent injection.

Distorted peak shapes can adversely affect quantitative determinations.

Baseline disturbances.

Variations in GC operating parameters.

Suggested Remedy

Develop a reproducible injection technique. Use autosampler or replace injection needle.

Correct any problems that result in the distortion of peak shape.

Disturbances in baseline are affecting peaks. ne problems.

Standardize parameters.

Negative Peaks Some or all peaks dip below the baseline.

Symptom







Possible Cause

Detector overload in elementspecific detectors such as ECD, NPD, FPD, etc. can produce both positive and negative peaks.

Dirty ECD detector can give a negative peak after a positive

Sample contaminants (hydrocarbons or other non-responders) are present when using ECD, PID, or NPD (thermoionic specific) detectors.

Incorrect polarity of the recorder connections results in nearly all peaks being negative.

Recorder-integrator wires reversed.

Sample injected onto the wrong column for dual-column setups.

Detector contamination.

Sample contamination.

Often normal for NPD (thermoionic specific) detectors.

Suggested Remedy

Have the compounds of interest arrive at the detector at a different time from the solvent or other compounds in high concentration. H₂ produces negative peaks with a TCD and helium carrier gas.

Clean or replace the ECD detector.

See detector maintenance.

Improve sample preparation and cleanup methods prior to injection.

Reverse polarity of recorder connections.

Correct connections.

Reinject the sample onto the correct column.

Clean or bake out the detector.

For PID detectors, check that the sample has not been contaminated with methanol or water. If necessary, prepare a fresh sample.

No correction necessary.

PEAK SHAPE PROBLEMS

No Peaks Some or all peaks are missing from the run.

Symptom



Possible Cause

Defective or clogged syringe.

"Blown" septum or massive leaks at the inlet.

Problems with carrier gas flow.

Column may be broken or installed in the wrong detector

The detector is not functioning or is not connected to the recorder or integrator.

Incorrect injector temperature: • Injector too cold: sample is not vaporized.

• Injector too hot: thermally labile sample is decomposing.

No Peaks After Solvent Peak



Sample volume is too high.

For FID detectors, the flame is blown out by the solvent peak.

Carrier gas flow is too high.

Incorrect column temperature; column is too hot and sample is eluting in solvent peak.

Column cannot separate components from solvent.

Activity in the inlet liner or column if the missing peak is an active compound.

Suggested Remedy

Try a new or proven syringe.

Find and fix leaks

Adjust gas flow.

If breakage is close to the beginning or end, cut off the short piece. Breakage in the middle can be repaired with a press-fit connector. For multiple breakages, replace or reinstall the column.

See column installation.

Ensure detector is working properly. (e.g. is the flame in a FID lit?) Check connection to the output device.

Cold injector: check injector and oven temperature with an accurate thermometer. If accurate, increase temperature as necessary, but do not exceed the maximum temperature limit of the column. Inject the sample directly onto the column.

Hot injector: check injector and oven temperature with an accurate thermometer. If accurate, reduce temperature as necessary, ensuring compatibility with sample and column minimum limit.

Inject less sample or use a higher split ratio

Check the detector temperature.

Reduce the flow rate.

Check oven temperature with an accurate thermometer. If accurate, reduce temperature as necessary, ensuring compatibility with sample and column minimum limit.

Change solvent or column.

Clean or replace the inlet liner. Ensure an inert column is used. If necessary, replace the column. See column installation.

Some Peaks Missing

Peaks Added There are more peaks than normal in the run.

Symptom





Possible Cause

Septum bleed (especially for runs with oven ramp).

Carryover of sample or contaminants from previous runs.

Contaminants in current sample or solvent.

Impurities in carrier gas are eluting.

Analytes are decomposing or breaking down for active or thermally labile compounds.

Suggested Remedy

Turn off the injector heater. If extra peaks disappear, choose a higher temperature rated septum or use a lower injection temperature.

Increase the analysis time prior to the next run or bake out the column between runs.

Inject solvent by itself using a clean syringe. Switch to a higher quality solvent if extra peaks appear. If only solvent appears, run the solvent through any sample preparation methods, analyzing the solvent at each step of the process to identify the source of extra peaks. If only the solvent peak appears, the extra peaks are part of the sample.

Install or check gas purifiers. Replace if necessary. Ensure only high-quality gases are used.

If compounds are thermally labile, lower the temperature and use on-column injection, a column with thinner stationary phase, a shorter column lengths, or a higher carrier gas flow rate.

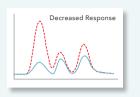
If compounds are active, ensure an inert column is used. If necessary, replace the column. See column installation



PEAK SHAPE PROBLEMS

Sensitivity Loss Some or all peaks are displaying decreased response.

Symptom



Possible Cause

Contamination of column and/or liner can lead to loss of sensitivity for active compounds.

Injector leaks reduce the peak height of the most volatile components of a sample.

Initial column temperature is too high for splitless injection prevents refocusing of sample. This affects the more volatile components most.

Inlet discrimination: injector temperature is too low. Later eluting and less volatile compounds have low response.

Issues with the sample.

Suggested Remedy

Clean liner.

See injector maintenance.

Bake out the column. Solvent rinse or replace the column. See column installation

Find and fix any leaks.

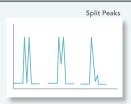
Lower temperature below the boiling point of the solvent. Decrease the initial column temperature, or use a less volatile

Increase the injection temperature or use on-column injection with direct connect liner.

Check the sample concentration, any sample preparation procedures, and shelf life. Prepare a fresh sample ensuring the proper concentration.

Split Peaks Peaks are duplicated or separated.

Symptom



Possible Cause

Poor (jerky or erratic) injection for manual injection.

Bad column installation.

Solvent mismatch: polarity of the stationary phase does not match the polarity of the solvent.

Wrong inlet liner is not vaporizing samples in one location.

Fluctuations in column temperature.

Mixed sample solvent for splitless or on-column injections.

Improper use of "solvent effect" refocusing techniques result in broad, distorted peaks because solutes are not refocused into a narrow band near the beginning of the column. The solvent must form a compact, continuous flooded zone in the column. If the solvent does not wet the stationary phase sufficiently (as might be the case for methanol used with a nonpolar phase), the flood zone may be several meters long and not of uniform thickness.

Suggested Remedy

Use smooth, steady plunger depression. Use autosampler.

Reinstall column. See column installation.

Change solvents, use a very large split ratio, install a retention gap, or change the stationary phase.

Use a liner with glass wool in the middle of the liner if possible.

Repair temperature control system.

Use a single solvent.

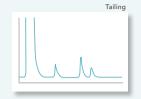
Install a retention gap (5 meters of uncoated but deactivated column) ahead of the column to reduce or eliminate the problem.

Change solvent or GC column

Use a very high split ratio.

Tailing Moderate to severe asymmetry towards the back or right side of the peak.

Symptom



Possible Cause

Contaminated inlet liner or column.

Activity in the inlet liner or column if the missing peak is an active compound.

Dead volume due to poorly installed liner or column.

Ragged column end.

Solvent-phase mismatch.

A cold region in the sample flow path.

Column or inlet liner temperature is too low for tailing hydrocarbons.

Debris in the liner or column.

Injection takes too long.

Split ratio is too low.

Overloading the inlet.

Some types of compounds such as alcoholic amines, primary and secondary amines, and carboxylic acids tend to tail.

Suggested Remedy

Clean or replace inlet liner. Bake out or replace the column.

Clean or replace the inlet liner. Ensure an inert column is used. If necessary, replace the column.

Confirm by injecting inert peak methane; if it tails, column is not properly installed. Reinstall liner and column as necessary.

Score the tubing lightly with a ceramic scoring wafer before breaking it. Examine the end (a 20-power magnifying glass is recommended). If the break is not clean and the end square, cut the column again. Point the end down while breaking it, and while installing a nut and ferrule. to prevent fragments from entering the column. Reinstall the column.

See column installation.

Change the stationary phase. Usually polar analytes tail on non-polar columns, or dirty columns.

Remove any cold zones in the flow path or check the MS transfer line trap.

Check injector and oven temperature with an accurate thermometer. If accurate, increase temperature as necessary, but do not exceed the maximum temperature limit of the column.

Clean or replace the liner. Cut 4 inches off the end of the column and reinstall it.

See column installation.

Improve injection technique.

Increase split ratio to at least 20:1.

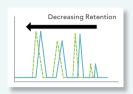
Decrease the sample volume or dilute the sample

Try a more polar column. Derivatize the sample.

PEAK SHAPE PROBLEMS

Retention Time Shifts Peak retention times drift or move.

Symptom



Increasing Retention

Irreproducible Retention

Possible Cause

Increase in column temperature.

Increase in gas flow rate (linear velocity).

Change of solvent.

Significant loss of stationary phase due to column bleed.

Leak in the injector.

Decrease in column temperature.

Decrease in gas flow rate (linear velocity).

Poor (jerky or erratic) injection for manual injection.

Contaminated column.

Leak in the injector.

Near-empty carrier gas tank.

Suggested Remedy

Check GC oven temperature and adjust as necessary. Ensure run conditions do not exceed the minimum temperature limits of the column.

Inject a detectable unretained sample such as methane to determine the linear gas velocity. Adjust gas pressure to obtain proper values for your analytical method.

Use the same solvent for standards and samples.

Reduce oven temperature. Ensure run conditions do not exceed the maximum temperature limit of the column.

Replace the column if necessary. See column installation.

Find the leak and fix it. Check the septum first. Change if necessary.

Check GC oven temperature and adjust as necessary. Ensure run conditions do not exceed the maximum temperature limits of the column.

Inject a detectable unretained sample such as methane to determine the linear gas velocity. Adjust gas pressure to obtain proper values for your analytical method.

Use smooth, steady plunger depression. Use autosampler.

Bake out the column. Cut 4 inches off the end of the column. Solvent rinse or replace the column.

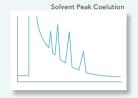
See column installation.

Find the leak and fix it. Check the septum first. Change if necessary.

Check and replace the tank if necessary.

Solvent Peak Broad The solvent peak is wide and coeluting with analyte peaks.

Symptom



Possible Cause

Bad column installation.

Injector leak.

Injection volume too large.

Injection temperature too low.

Split ratio is too low.

Column temperature too low.

Initial column temperature too high for splitless injection.

Purge time (splitless hold time) too long for splitless injection.

Suggested Remedy

Reinstall column.

Find and fix leak

Decrease sample injection volume or dilute to 1:10.

Increase injection temperature so the entire sample is vaporized "instantly." An injection temperature higher than the temperature limit of the column will not damage the column.

Increase split ratio.

Increase column temperature. Use a lower boiling solvent.

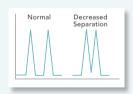
Decrease the initial column temperature.

Use a less volatile solvent so the initial column temperature is below the boiling point of the solvent.

Use a shorter purge valve closed

Resolution Loss Peaks begin to coelute or overlap.

Symptom



Possible Cause

Change in column dimensions or stationary phase; excessive column trimming.

Damage to column stationary phase.

Damage to column stationary

Injector problems.

phase.

Suggested Remedy

Differences in retention time or peak shape of other compounds will be apparent. Check the column phase and dimensions switch the column if necessary.

This is usually indicated by excessive column bleed. Replace the column.

See column installation.

This is usually indicated by excessive column bleed. Replace the column.

See column installation.

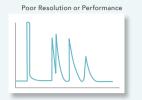
Check for:

- leaks
- inappropriate temperature
- split ratio
- purge time
- dirty liner
- glass wool in liner

PEAK SHAPE PROBLEMS

Performance Loss (Column) The column deteriorates too quickly after installation.

Symptom



Possible Cause

Column too hot for too long

Exposure to oxygen, particularly at elevated temperatures.

Chemical damage due to inorganic acids or bases.

Contamination of the column with nonvolatile materials.

Broken column.

Suggested Remedy

Stay below limits specified for the column. Replace column.

Find and fix any leaks. Be sure carrier gas is sufficiently pure.

Keep inorganic acids or bases out of column. Neutralize samples

Find and fix any leaks. Be sure carrier gas is sufficiently pure.

Keep inorganic acids or bases out of column. Neutralize samples.

Prevent nonvolatile materials from getting into column. For example, use a quard column or a Zebron[™] column with Guardian™ integrated guard.

Replace column. See column installation.

Avoid damaging the polyimide coating on the column. Except when using Zebron Inferno™ GC columns, avoid temperatures above 370 °C; abrasion of columns (for example, do not install a column so that it touches the side of the oven, because vibration may then damage the polyimide coating); or excessive bending or twisting, which will damage this protective coating. Remember, even if the column does not break immediately, when the protective coating is damaged the column may break spontaneously later.





A Note On Solvent Rinsing

final rinse if you are using an ECD. Avoid acetonitrile as a final rinse if you are using an NPD. Methanol,

Each solvent should remain in the column for at least 10 minutes. There is no need to remove the prebe purged with pure carrier gas for 10 minutes before reinstallation. Program the oven temperature at

COLUMN SELECTION PROBLEMS

The Impact of Selectivity

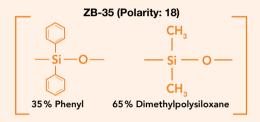
Resolution between two analytes is mainly determined by the selectivity of the stationary phase. By increasing the resolution between two compounds, the total analysis time can often be reduced significantly!

The Master Resolution Equation

$$R_{s} = \begin{bmatrix} \sqrt{N} \\ 4 \end{bmatrix} \times \begin{bmatrix} \alpha - 1 \\ \alpha \end{bmatrix} \times \begin{bmatrix} \frac{k}{k+1} \end{bmatrix}$$
Efficiency Term Selectivity Term Retention Term

Selectivity vs. Polarity

Column polarity and selectivity are often confused – polarity gives a general guideline for sample capacity and separation, which can affect peak shape and resolution. However, two columns may have similar polarity but show very different separation profiles because of differences in phase chemistry. Example: The ZB-1701 cyanopropyl group makes it very different from ZB-35 in terms of selectivity though polarity is similar.



The 3 Most Prevalent GC Interactions

Dispersive Forces (Van der Waals Interactions)

- Weakest of all intermolecular forces and occurs between non-polar compounds
- Separation is based on boiling point (classic example hydrocarbon separation in simulated distillation analysis)

Dipole-Dipole Interactions

- Either permanently present or induced by analyte-stationary phase interactions
- Higher dipole-dipole interaction can help separate compounds with similar boiling points, but different chemical structures

Hydrogen Bonding (Acid-Base Interactions)

- Can cause poor peak shape or irreversible binding to the inlet liner or to the column itself
- Zebron[™] columns are specially deactivated to minimize these interactions

Dimension Selection

COLUMN SELECTION PROBLEMS

Dimension Selection

Short

(15 m or less)

Applications

- High boilers
- GC-MS applications

Advantages

- Faster run times
- Higher temp. limits

· Higher efficiency

Lower bleed

Disadvantages

- Less inert
- Limited retention



Applications

- Complex samples with closely eluting peaks
- Low boilers
- Less active samples
- Complex temperature ramps

Advantages

Better resolution

Disadvantages

Long

(60m or more)

Slow run times

Narrow

(0.10, 0.18, or 0.20 mm)

Applications

Complex samples

Advantages

- Faster run times
- Better resolution

0.25mm



INTERNAL DIAMETER

Applications

- Dirty samples
- Highly concentrated samples

Advantages

- Increased sample capacity
- Good for routine on-column injection

- **Disadvantages**
- Decreased efficiency
- May need higher flow rates Not compatible with most GC-MS

Thick

(0.50 µm or more)

Wide

(0.32 or 0.53 mm)

Thin

(0.10 or 0.18 µm)

Applications

- High boilers
- GC-MS applications

Advantages

- Faster run times
- Higher temp. limits Lower bleed Higher efficiency

Disadvantages

Disadvantages • Lower sample capacity

• Easily overloaded

- Less inert
- Limited retention

FILM THICKNESS 0.25 µm

Applications

- Low boilers
- Gases, solvents, purgeables, volatiles
- Purity testing

Advantages

- Better inertness
- Higher capacity

Disadvantages

- Slower run times
- Lower temp. limits
- Higher bleed

Phase Selection Chart

Content	Polarity	Phase	Composition	Temperature Limits (Isothermal/TPGC)	GC-MS Certified	Recommended Applications
100 100	Essentials Non-Polar	ZB-1	100% Dimethylpolysiloxane	-60 to 360/370°C*	✓	
Section Sect	Non-Polar	ZB-5		-60 to 360/370°C*	/	Alkaloids, Dioxins, Drugs, Essential Oils/Flavors, Halo-hydrocarbons, PCBs/Aroclors, Pesticides/Herbicides, Phenols,
Section Sect	Non-Polar	ZB-5ms		-60 to 325/350°C	✓	
Solventry Solv	Intermediate	ZB-35		40 to 340/360°C	1	Amines, Aroclors, Drugs, EPA Methods (508, 608, 8081, 8141, 8151), Pesticides, Pharmaceuticals, Semi-volatiles, Steroids
Marchella	Intermediate	ZB-624		-20 to 260°C		
Transplations 78-50 50 Simular phylopholisman 78-50 78	Intermediate	ZB-XLB	Proprietary	30 to 340/360°C*	✓	PCBs, Pesticides/Herbicides
1906 20 MAX 100 Foperhyland (20 to 20 to 2000 C) 100 Foperhyland (20 to 20 t	Intermediate	ZB-1701		-20 to 280/300°C*		
Soverts, System, Alymon Exement Soverts, System, System Exement Soverts, System, System Exement Soverts, System, Alymon Exemption, Figure Soverts, System, Alymon Exemption, Special Call Soverts, Special Call Soverts, System, Alymon Exemption, Special Call Soverts, System, Alymon Exemption, Special Call Soverts, System, Alymon Exemption, Figure Sove	Polar	ZB-50		40 to 320/340°C*	✓	
The Properties of the Control of the	Polar	ZB-WAX	100% Polyethylene Glycol	20 to 250/260°C	✓	
Non-Polar ZB-HTT 100% Dimethylpolyslovane 46 to 400/430°C / CPRCs, High Bolling Persolaum Products, High Molecular Weight Wases, Long-chained Hydrocarbons, Motor Olls, Phylophylpolysion 25 to Phylop	Polar	ZB-FFAP		40 to 250/260°C		Acrylates, Alcohols, Aldehydes, Free Fatty Acids, Ketones, Organic Acids, Phenols, Volatile Free Acids
Non-Polar 28-91T (10% Directlyphysiological 40 to 400/430°C / Dissiplication Dissiplication State Products, Significant Dissiplication State Products, Significant Dissiplication, Surface across, Fighycerides 5% Picesy 5% Directlyphysiological 5% Picesy 35% Picesy 100% Directlyphysiological 23.5 Memory 100% Directlyphysiological 23.5 Memory 100% Directlyphysiological 23.5 Memory 100% Directlyphysiological 24.0 Memory 100% Directlyphysiological 25.5 Memory 100% Directlyphysiological	Inferno™					
Histornediate 28.35HT 65% Dimethylophylopaloxane 35% Phenyl Phylophylopaloxane 35% Phenyl Phylophylophylopaloxane 35% Phenyl Phylophylophylophylophylophylophylophylop	Non-Polar	ZB-1HT	100% Dimethylpolysiloxane	-60 to 400/430°C*	✓	
recording the seconds of the second o	Non-Polar	ZB-5HT		-60 to 400/430°C*	✓	
Non-Polar ZB-1rus** 100% Dimethylpolysiloxane 40 to 300/30°C* / Acids, Amines, Diesel Fuel, Drugs, Flavors & Fragrances, PCBs (EPA Method 1668), Pesticides Non-Polar ZB-5rus** 9% Dimethylpolysiloxane 5% Phenys 5% Phe	Intermediate	ZB-35HT		40 to 400°C	✓	
Non-Polar ZB-1Pus" 100% Dimethylpolysiloxane 4.00 to 360/370°C / Acids, Amines, Diesel Fuel, Drugs, Flavors & Fragrances, PCBs (EFA Method 1668), Pesticides Non-Polar ZB-5Pus" 5% Dimethylpolysiloxane 5% Phenol 3% Phenol 5% Phe	Intermediate	ZB-XLB-HT	Proprietary	30 to 400°C	√	EPA Methods, PCBs, Pesticides/Herbicides
Non-Polar ZB-Srius** 95% Dimethylpolysiloxane 5% Phenyl 40 to 326/370°C / Drugs, EPA Methods, Nitrosamines, Pesticides, Phenols 5% Phenyl 40 to 326/350°C / Britishylpolysiloxane 5% Dimethylpolysiloxane 5% Dimethylpolysiloxane 5% Dimethylpolysiloxane 5% Phenyl 40 to 320/320°C / Canabis, Terpenes, Residual Solvents, Volatile Amines, EPA Methods (524, 624, 8260), Food, Flavors, Fragrances, Solvent Purity, Alcohols 28-WAX.rius** 100% Polyethylene Glycol 20 to 250/260°C* Canabis, Terpenes, Residual Solvents, Volatile Amines, EPA Methods (524, 624, 8260), Food, Flavors, Fragrances, Solvent Purity, Alcohols Alcohols, Aldelydes, Aramatics, Essential Oils, Flavors & Fragrances, Glycols, OVIs, Pharmaceuticals, Solvents / Residual Solvents, Styrene, Xylene Isomers Non-Polar ZB-NXT SimDist 100% Dimethylpolysiloxane -60 to 450°C* / ASTM Methods (5288, D2887X, D3710, D6352, D7169), Crude Oil, Gasoline Fractions, Petroleum Distillates Non-Polar ZB-DHA-PONA 100% Dimethylpolysiloxane -60 to 360/370°C / DHA, PONA, PIANO, PASTM Methods (50154, D5441, D5501, D6729, D6730, D6733) Non-Polar ZB-Semivolatiles 5% Dimethylpolysiloxane -60 to 325/350°C / Semi-volatiles (SVOCs), PAHs, EPA Methods (528, 610, 625, 8100, 8270D) Intermediate ZB-MultiResidue**1 & 2 Proprietary -60 to 320/340°C / Dal-Colombine Petricides, Organophopiprorus Petricides, Organophopiprorus Petricides, Organophopiprorus Petricides, Organophopiprorus Petricides, Organophopiprorus Petricides, Polar Proprietary -60 to 340/360°C / Dual-Column Chlorinated Petracides by G-EC-DE, PAM Ethods (8081, 8082, 8151, 504, 505, 508, and 552) Proprietary ZB-BAME High Cyanopropyl -01 to 260/280°C / Daus-Column Chlorinated Petracides by G-EC-DE, PAM Ethods (8081, 8082, 8151, 504, 505, 508, and 552) Proprietary ZB-Boushanol Proprietary -60 to 340/360°C / Drug Screening (6-MAM, Amphetamines, Barbiturates, Benzodiazepines, Opistes, PCP, THC) Proprietary ZB-Boushanol Proprietary -60 to 340/360°C / Analysis of 15+1 EU-Regulated and EPA-Regulated PAHs in Food Testing, Rubber, Plastic, Coal	PLUS					
Non-Polar ZB-SMS-Pilus" 9% Blomethylopisiloxane 5% Phenyl-Arylene 40 to 325/350°C / Acids, Aladisids, Amines, Drugs, Ethanolamines, Essential Oiis/Flavors, Halo-hydrocarbons, Pesticides/Herbicides, Phenols, Solvent Impurities Non-Polar ZB-SMS-Pilus" 100% Polyethylene Glycol 20 to 250/260°C / Cannabis, Terpenes, Residual Solvents, Volatile Amines, EPA Methods (524, 624, 8260), Food, Flavors, Fragrances, Solvent Putty, Alcohols Non-Polar ZB-MXF. 100% Polyethylene Glycol 20 to 250/260°C / Alcohols, Aldehydes, Aromatics, Essential Oils, Flavors & Fragrances, Glycols, OVIs, Pharmaceuticals, Solvents / Residual Solvents / Syrene, Xylene Isomers ***Unlimited*** ***Unlimited*** ***Non-Polar ZB-MXT SimDist 100% Dimethylpolysiloxane -60 to 450°C / ASTM Methods (D2887, D2887X, D3710, D4532, D7169), Crude Oil, Gasoline Fractions, Petroleum Distillates, Petroleum Practions, Simulated Distillation, Vacuum Distillates ***Non-Polar ZB-DHA-PONA 100% Dimethylpolysiloxane -60 to 360/370°C / DHA, PONA, PIONA, PIONA, PIONA, DATM Methods (D254, D541, D5501, D6729, D6730, D6730, D6733) Non-Polar ZB-SemiVolatiles 5% Dimethylpolysiloxane -60 to 320/340°C / DHA, PONA, PIONA, PIONA, PIONA, PIONA, D452, D544, D5501, D6729, D6730, D6730	Non-Polar	ZB-1 <i>PLUS</i> ™		-60 to 360/370°C*	✓	Acids, Amines, Diesel Fuel, Drugs, Flavors & Fragrances, PCBs (EPA Method 1668), Pesticides
Phenols, Solvent Impurities Intermediate Z8-64#,US* Proprietary -20 to 300/320*C / Canabis, Teppense, Residual Solvents, Volatile Amines, EPA Methods (524, 624, 8260), Food, Flavors, Fragrances, Solvent Purity, Alcohols, Aldehydes, Aromatics, Essential Oils, Flavors & Fragrances, Glycols, OVIs, Pharmaceuticals, Solvents, Syrene, Nyelene Isomers ***Unlimited*** ***Non-Polar** Z8-IXT SimDist*** ***Non-Polar** Z8-IXT SimDist*** ***100% Dimethylpolysiloxane*** -60 to 450*C*** ***50 The Apylor Color Colo	Non-Polar	ZB-5 <i>PLUS</i> ™	5% Phenyl	-60 to 360/370°C	✓	
Polar ZB-WAXPLUS** 100% Polyethylene Glycol 20 to 250/260°C* Alcohols, Aldehydes, Aromatics, Essential Oils, Flavors & Fragrances, Glycols, OVIs, Pharmaceuticals, Solvents / Residual Solvents, Styrene, Xylene Isomers Won-Polar ZB-IXT SimDist 100% Dimethylipolysiloxane -60 to 450°C* / ASTM Methods (D2887, D2887X, D3710, D6352, D7169), Crude Oil, Gasoline Fractions, Petroleum Distillates, Petroleum Fractions, Simulated Distillation, Vacuum Distillates Non-Polar ZB-SemiVolatiles 95% Dimethylipolysiloxane -60 to 360/370°C / DHA, PONA, PIONA, PIANO, ASTM Methods (D5134, D5441, D5501, D6729, D6730, D6733) Non-Polar ZB-SemiVolatiles 95% Dimethylipolysiloxane -60 to 320/340°C / Semi-volatiles (SVOCs), PAHs, EPA Methods (S25, 610, 625, 8100, 8270D) Intermediate ZB-MultiResidue*-1 &-2 Proprietary -60 to 320/340°C / Aroclors/PCBs, Haloacetic Acids, Herbicides, Insecticides, Multi-Pesticide Screening, Nitrogen Containing Pesticides, Organochhorine Pesticides, Organophosphorous Pesticides ZB-FAME High Cyanopropyl -20 to 280°C / Dual-Column Chlorinated Pesticides by GC-ECD, EPA Methods (8081, 8082, 8151, 504, 505, 508, and 552) Proprietary ZB-BAC-1 &-2 Proprietary -20 to 260/280°C / Dual-Column Chlorinated Pesticides by GC-ECD, EPA Methods (8081, 8082, 8151, 504, 505, 508, and 552) Proprietary ZB-Bochanol Proprietary -60 to 340/360°C / Drug Screening (6-MAM, Amphetamines, Barbiturates, Benzodiazepines, Opiates, PCP, THC) Proprietary ZB-Bioethanol Proprietary -60 to 340/360°C / Dioxin and PCB in Food and Environmental Samples, POPs in Food Proprietary ZB-Dioxin Proprietary -40 to 320/340°C / Dioxin and PCB in Food and Environmental Samples, POPs in Food Analysis of 15+1 EU-Regulated and EPA-Regulated PAHs in Food Testing, Rubber, Plastic, Coal	Non-Polar	ZB-5MS <i>PLUS</i> ™		-60 to 325/350°C	✓	Phenols, Solvent Impurities
Unlimited Von-Polar ZB-TXT SimDist 100% Dimethylpolysiloxane -60 to 450°C* / ASTM Methods (D2887, D2887X, D3710, D6352, D7169), Crude Oil, Gasoline Fractions, Petroleum Distillates, Petroleum Distillates, Petroleum Practicions, Simulated Distillation, Vacuum Distillates Non-Polar ZB-DHA-PONA 100% Dimethylsiloxane -60 to 360′370°C / DHA, PONA, PIONA, PIANO, ASTM Methods (D5134, D5441, D5501, D6729, D6730, D6733) Non-Polar ZB-SemiVolatiles 95% Dimethylpolysiloxane 55% Phenyi-Arylene -60 to 320′340°C / Semi-volatiles (SVOCs), PAHs, EPA Methods (525, 610, 625, 8100, 8270D) Intermediate ZB-MultiResidue*1.8 -2 Proprietary -60 to 320′340°C / Aroclors/PCBs, Haloacetic Acids, Herbicides, Insecticides, Multi-Pesticide Screening, Nitrogen Containing Pesticides, Organophorphorous Pesticides in Periodic Proprietary ZB-BAC-1.8 -2 Proprietary -20 to 260′280°C / Dual-Column Chlorinated Pesticides by GC-ECD, EPA Methods (8081, 8082, 8151, 504, 505, 508, and 552) Proprietary ZB-Drug-1 Proprietary -20 to 260′280°C / Drug Screening (S-MAM, Amphetamines, Barbiturates, Benzodiazepines, Opiates, PCP, THC) Proprietary ZB-Dioxin Proprietary -40 to 320′340°C / Drug Screening (S-MAM, Amphetamines, Barbiturates, Benzodiazepines, Opiates, PCP, THC) Proprietary ZB-Dioxin Proprietary -40 to 320′340°C / Dioxin and PCB in Food and Environmental Samples, POPs in Food Proprietary -40 to 340′360°C / Analysis of 15+1 EU-Regulated and EPA-Regulated PAHs in Food Testing, Rubber, Plastic, Coal	Intermediate	ZB-624PLUS™	Proprietary	-20 to 300/320°C	✓	Purity, Alcohols
Non-Polar ZB-1XT SimDist 100% Dimethylpolysiloxane -60 to 450°C* / ASTM Methods (D2887, D2887X, D3710, D6352, D7169), Crude Oil, Gasoline Fractions, Petroleum Distillates, Petroleum Practions, Simulated Distillation, Vacuum Distillates Non-Polar ZB-DHA-PONA 100% Dimethylsiloxane -60 to 360/370°C / DHA, PONA, PIONA, PIONA, PIONA, PIONA, PIONA, DSTM Methods (D5134, D5411, D5501, D6729, D6730, D6733) Non-Polar ZB-SemiVolatiles SS-SemiVolatiles ZB-MultiResidue"-1 & -2 Proprietary 2B-BAC-1 & -2 Proprietary ZB-Dixin Proprietary A0 to 320/340°C / Dixin and PCB in Food and Environmental Samples, POPs in Food Posting, Rubber, Plastic, Coal	Polar	ZB-WAX <i>PLUS</i> ™	100% Polyethylene Glycol	20 to 250/260°C*		
Non-Polar ZB-DHA-PONA 100% Dimethylpolysiloxane -60 to 360/370 °C / DHA, PONA, PIONA,	Unlimited					
Non-Polar ZB-SemiVolatiles 95% Dimethylpolysiloxane 5% Phenyl-Arylene -60 to 325/350 °C -60 to 325/350	Non-Polar	ZB-1XT SimDist	100% Dimethylpolysiloxane	-60 to 450°C*	1	
Intermediate ZB-MultiResidue™-1 & -2 Proprietary -60 to 320/340°C ✓ Aroclors/PCBs, Haloacetic Acids, Herbicides, Insecticides, Multi-Pesticide Screening, Nitrogen Containing Pesticides, Organochlorine Pesticides, Organophosphorous Pesticides Described Pesticides Described Pesticides Described Pesticides Described Pesticides Described	Non-Polar	ZB-DHA-PONA	100% Dimethylsiloxane	-60 to 360/370°C	✓	DHA, PONA, PIONA, PIANO, ASTM Methods (D5134, D5441, D5501, D6729, D6730, D6733)
Intermediate ZB-CLPesticides-1 & -2 Proprietary 40 to 320/340 °C / Dual-Column Chlorinated Pesticides by GC-ECD, EPA Methods (8081, 8082, 8151, 504, 505, 508, and 552) Polar ZB-FAME High Cyanopropyl -20 to 280 °C / Fatty acid methyl esters (FAMEs), Cis/trans FAME isomers Proprietary ZB-BAC-1 & -2 Proprietary 40 to 320/340 °C / Abused Inhalant Anesthetics, Blood Alcohol Analysis Proprietary ZB-Bioethanol Proprietary 40 to 320/340 °C / Alcohol, Bioethanol, Fusel Alcohols Proprietary ZB-Dioxin Proprietary 40 to 320/340 °C / Dioxin and PCB in Food and Environmental Samples, POPs in Food Proprietary ZB-PAH-EU Proprietary 40 to 340/360 °C / Analysis of 15+1 EU-Regulated and EPA-Regulated PAHs in Food Testing, Rubber, Plastic, Coal	Non-Polar	ZB-SemiVolatiles		-60 to 325/350°C	✓	Semi-volatiles (SVOCs), PAHs, EPA Methods (525, 610, 625, 8100, 8270D)
Polar ZB-FAME High Cyanopropyl -20 to 280 °C	Intermediate	ZB-MultiResidue [™] -1 & -2	Proprietary	-60 to 320/340°C	✓	
Proprietary ZB-BAC-1 & -2 Proprietary -20 to 260/280°C	Intermediate	ZB-CLPesticides-1 & -2	Proprietary	40 to 320/340°C	✓	Dual-Column Chlorinated Pesticides by GC-ECD, EPA Methods (8081, 8082, 8151, 504, 505, 508, and 552)
Proprietary ZB-Drug-1 Proprietary 40 to 320/340 °C	Polar	ZB-FAME	High Cyanopropyl	-20 to 280°C	✓	Fatty acid methyl esters (FAMEs), Cis/trans FAME isomers
Proprietary ZB-Bioethanol Proprietary -60 to 340/360°C ✓ Alcohol, Bioethanol, Fusel Alcohols Proprietary ZB-Dioxin Proprietary 40 to 320/340°C ✓ Dioxin and PCB in Food and Environmental Samples, POPs in Food Proprietary ZB-PAH-EU Proprietary 40 to 340/360°C ✓ Analysis of 15+1 EU-Regulated and EPA-Regulated PAHs in Food Testing, Rubber, Plastic, Coal	Proprietary	ZB-BAC-1 & -2	Proprietary	-20 to 260/280°C	✓	Abused Inhalant Anesthetics, Blood Alcohol Analysis
Proprietary ZB-Dioxin Proprietary 40 to 320/340 °C ✓ Dioxin and PCB in Food and Environmental Samples, POPs in Food Proprietary ZB-PAH-EU Proprietary 40 to 340/360 °C ✓ Analysis of 15+1 EU-Regulated and EPA-Regulated PAHs in Food Testing, Rubber, Plastic, Coal	Proprietary	ZB-Drug-1	Proprietary	40 to 320/340°C	✓	Drug Screening (6-MAM, Amphetamines, Barbiturates, Benzodiazepines, Opiates, PCP, THC)
Proprietary ZB-PAH-EU Proprietary 40 to 340/360°C ✓ Analysis of 15+1 EU-Regulated and EPA-Regulated PAHs in Food Testing, Rubber, Plastic, Coal	Proprietary	ZB-Bioethanol	Proprietary	-60 to 340/360°C	✓	Alcohol, Bioethanol, Fusel Alcohols
	Proprietary	ZB-Dioxin	Proprietary	40 to 320/340°C	✓	Dioxin and PCB in Food and Environmental Samples, POPs in Food
Proprietary ZB-PAH-CT Proprietary 40 to 320/340 °C 🗸 Analysis of 15+1 EU-Regulated and EPA- Regulated PAHs in Food Testing, Rubber, Plastic, Coal	Proprietary	ZB-PAH-EU	Proprietary	40 to 340/360°C	✓	Analysis of 15+1 EU-Regulated and EPA-Regulated PAHs in Food Testing, Rubber, Plastic, Coal
	Proprietary	ZB-PAH-CT	Proprietary	40 to 320/340°C	1	Analysis of 15+1 EU-Regulated and EPA- Regulated PAHs in Food Testing, Rubber, Plastic, Coal

GC Column Cross-Reference Chart

Zebron™ Phase	Restek®	Agilent® Technologies	Supelco®	Alltech®	SGE®	OV®
ZB-1	Rtx [®] -1, Rtx-1PONA, Rtx-1 F&F	DB®-1, DB-2887, DB-1 EVDX, HP-1, HP-101, HP-PONA, Ultra 1, CP-Sil 5 CB	SPB®-1, SPB-1 TG, SE-30, MET-1, SPB-1 Sulfur, SPB-HAP	AT [™] -1, AT-Sulfur, EC-1	BP1, BP1-PONA, BPX1-SimD	OV-1
ZB-DHA-PONA	Rtx-DHA	HP-PONA, DB-PETRO, CP-Sil PONA CB	Petrocol®-DH			
ZB-1₽LUS [™]	Rtx-1ms, Rxi®-1ms	DB-1ms, DB-1ms Ultra Inert, HP-1ms, HP-1ms Ultra Inert, CP-Sil 5 CB MS, VF-1ms	MDN™-1, Equity®-1	AT-1ms	SolGEL-1ms [™]	
ZB-1HT Inferno™	Rxi®-1HT	DB-1ht, CP-SimDist	Petrocol® 2887			
ZB-1XT SimDist	MXT®-1HT SimDist MXT-1, MXT-1 SimDist, MXT-2887	CP-SimDist UltiMetal, CP-Sil 8 CB UltiMetal, BPX1-SimD, DB-PS1, DB-HT SimDis, DB-PS2887				
ZB-5	Rtx-5	DB-5, HP-5, Ultra 2, HP-PAS-5, CP-Sil 8 CB, HP-5ms Ultra Inert	MDN-5, SPB-5, PTE-5, SE-54, PTA-5, Equity-5, Sac-5	AT-5, EC-5	BP5, BPX5	OV-5
ZB-5HT Inferno	Stx®-5HT, Rtx-5HT, Rxi-5HT, XTI®-5HT	DB-5ht, VF-5ht	HT-5			
ZB-5ms	Rtx-5Sil MS, Rxi-5Sil MS	DB-5ms, DB-5.625, DB-5ms EVDX, VF-5ms, CP-Sil 8 CB MS				
ZB-5₽LUS [™]	Rtx-5ms, Rtx-5Amine, Rxi-5ms	DB-5, HP-5ms, HP-5msi	MDN-5S			
ZB-5MS _{PLUS} ™	Rxi-5Sil MS	DB-5ms Ultra Inert, HP-5ms Ultra Inert, DB-5ms, VF-5ms	SLB®-5ms			
ZB-SemiVolatiles	Rxi-5Sil MS Rxi-5ms	DB-5ms Ultra Inert	SLB®-5ms			
ZB-35	Rtx-35, Rtx-35ms	DB-35, DB-35ms, HP-35, HP-35ms	MDN-35, SPB-35, SPB-608	AT-35	BPX35, BPX608	OV-11
ZB-35HT Inferno			Phenomenex Exclusive			
ZB-50	Rtx-50	DB-17, DB-17HT, DB-17ms, DB-17 EVDX, HP-50+, CP-Sil 24 CB	SP-2250, SPB-17, SPB-50	AT-50	BPX50	OV-17
ZB-624	Rtx-1301, Rtx-624	DB-1301, DB-624, DB-VRX, HP-VOC, CP-1301, CP-Select 624 CB	SPB-1301, SPB-624	AT-624, AT-1301	BP624	OV- 624
ZB-624 PLUS TM	Rxi-624Sil MS	CP-Select 624 CB, DB-624UI Ultra Inert				
ZB-1701	Rtx-1701	DB-1701 , CP-Sil 19 CB	SPB-1701, Equity-1701	AT-1701	BP10	OV -1701
ZB-1701P		DB-1701P				
ZB-WAX	Rtx-WAX, Famewax [™] , Stabilwax®-DB	DB-WAXetr, HP-INNOWax, CP-Wax 57 CB	Met-Wax, Omegawax®	EC-Wax	SolGEL-WAX™	
ZB-WAX _{PLUS} ™	Stabilwax®	DB-WAX, CAM, HP-20M, Carbowax 20M, CP-Wax 52 CB	SUPELCOWAX® 10	AT-Wax, AT-AquaWax	BP20	Carbowax 20M
ZB-FAME		CP-Sil 88, HP-88, DB-23	SP-2560, SP-2330			
ZB-FFAP	Stabilwax-DA	DB-FFAP, HP-FFAP, CP-Wax 58 (FFAP) CB, CP-FFAP CB	Nukol™, SPB-1000	AT-1000, EC-1000	BP21	OV-351
ZB-MultiResidue™-1	Rtx-CLPesticides, Stx-CLPesticides					
ZB-MultiResidue-2	Rtx-CLPesticides2, Stx-CLPesticides2					
ZB-CLPesticides-1	Rtx-CLPesticides, Stx-CLPesticides					
ZB-CLPesticides-2	Rtx-CLPesticides2, Stx-CLPesticides2					
ZB-XLB	Rtx-XLB, Rxi-XLB	DB-XLB, VF-XMS	MDN-12			
ZB-XLB-HT Inferno			Phenomenex Exclusive			
ZB-Drug-1			Phenomenex Exclusive			
ZB-BAC1	Rtx-BAC-1	DB-ALC3				
ZB-BAC2	Rtx-BAC-2	DB-ALC2				
ZB-Bioethanol		Pau Diavia 2	Phenomenex Exclusive			
ZB-Dioxin	Dwi DALI	Rtx-Dioxin-2	SP-2330			
ZB-PAH-EU	Rxi-PAH	DB-PAH-EU				
ZB-PAH-CT		PAH-Select				

PROBLEM PREVENTION



PROBLEM PREVENTION

The Inlet: Sample Injection Techniques

One size does not fit all – there is no single injection mode that accommodates all samples and all columns. Instead, an appropriate injection mode introduces the sample so that it:

- Retains its original composition (i.e. there should be no sample degradation or selective losses during injection)
- Occupies the shortest possible length of column (the shorter the initial sample band, the sharper the peaks, the better the sensitivity, and the better the resolution)

Injection Modes: Split

In split injection, the sample is rapidly vaporized and mixed with carrier gas. Most of the sample is vented through the split vent, while a small amount enters the column. The flow through the split vent divided by the flow through the column is called the "split ratio". This rapid sample introduction provides the basis for sharp peaks and good resolution; it may however be inappropriate if sample components vary widely in their boiling points.

Inlet Discrimination | The less volatile components of a sample will not vaporize as rapidly, so immediately after injection the vaporized sample has a greater proportion of the more volatile compounds than the original sample. This effect is called "discrimination". The longer the sample spends in the heated inlet, the less the discrimination – but the broader the peaks.

Backflash | Backflash occurs when the vaporized sample expands and exceeds inlet liner volume. Vapors may come in contact with cold spots (e.g. the septum or inlets to the injector) and less volatile components may condense. These condensates may vaporize later and interfere with subsequent analyses, sometimes producing "ghost peaks." Expansion outside the liner may also expose the sample to active metal surfaces and reactive components of the sample may be lost. Minimized backflash by using a septum purge, small injection volumes, large volume inlet liners, and optimal inlet temperatures.

Inlet Temperature | The temperature should be hot enough to ensure rapid vaporization of the entire sample, but not too hot to degrade any analytes. Experimentation may be required to mitigate inlet discrimination and backflash. A good starting point is 250°C.

Septum Purge | Gas sweeping the bottom face of the septum and through a purge vent carries contaminants out. Higher than optimum purge flows may result in the loss of more volatile sample components. Septum purge flow rates are usually between 0.5 and 5 mL/min.

Sample Size & Concentration | Split injection is used for highly concentrated samples. Typical concentrations are from $0.1-10\,\mu g/\mu L$. Injection volumes of 1 to $2\,\mu L$ are common, and up to $5\,\mu L$ can be used without great problems, depending on the solvent used. If the sample volume is too large, backflash may occur.

Injection Modes: Splitless

In splitless injection, the entire flow through the injector passes into the column for the first 15 to 90 seconds, and is then refocused.

The Solvent Effect | To avoid the broad peaks that would otherwise result from slow split injections, samples are refocused before starting the chromatographic process following splitless injection. Refocusing can be accomplished by adjusting the initial column temperature to 10 °C or more below the boiling point of the sample solvent. When the vapor leaves the injector and enters the cooler column, the solvent condenses at the front of the column as a liquid band; vapors will condense in this band and be trapped and refocused. This process is called the "solvent effect." Improper use of solvent effect techniques result in broad, distorted peaks because solutes are not refocused into a narrow band near the beginning of the column. The solvent must form a compact, continuous flooded zone in the column. If the solvent does not wet the stationary phase sufficiently (as may be the case for methanol with a nonpolar phase), the flood zone may be several meters long and not of uniform thickness.

PROBLEM PREVENTION

The Inlet: Sample Injection Techniques

Cold Trapping | Solutes that boil at 150 °C or more above the initial column temperature do not require the solvent effect in order to refocus. These high boiling compounds will condense at the beginning of the column in a short band without the aid of the solvent. This process is called "cold trapping." Both the solvent effect and cold trapping can be achieved by operating in a temperature programmed mode.

Sample Volume | Samples are usually limited to $2\mu L$ or less to avoid overloading the inlet liner and the column. Sample injection volumes must be reproducible in order to obtain reproducible retention times or quantitative data.

Injection Modes: On-Column

On-column injection can eliminate syringe and inlet-related discrimination. If polar solvents are used with non-polar column linings, a retention gap is recommended. If the solvent is below boiling point at injection, the sample is distributed over a flooded zone at the front of the column and less volatile compounds are distributed in the phase. As the carrier gas evaporates the solvent at the front end, volatile components are concentrated and refocused.

Sample Focusing | The distribution of solutes in the area of the flooded zone is not homogeneous and this leads to peak broadening; this can be neglected for many applications and good quantitative results can still be obtained. If the compound boiling points are vastly different from the solvent, ballistic heating to high temperature can be employed. If compound boiling points are close to the solvent, temperature programming can be used. Wide-bore columns make on-column injection easier; alternatively a deactivated but uncoated wide-bore retention gap may be connected to a narrow-bore column.

Sample Size | Samples between 1 and $2\mu L$ can be injected rapidly into a column below the boiling point of the solvent. To keep the flooded zone short, sample size should be limited to $1\mu L$.

Injection Modes: Direct Injection

Direct injection should not be confused with on-column injection. It is a flash vaporizing method in which the inlet system is heated independently from the column oven. Sample evaporation occurs in the inlet.

Injection Modes: Programmed Temperature Vaporizing (PTV)

In PTV injection, the liquid sample is injected into a cold glass liner. After withdrawal of the syringe needle, the vaporizing tube is heated in a controlled manner (usually rapidly) to vaporize the sample. This injection method permits special handling of the sample to vent the solvent, or to avoid thermal decomposition of thermally labile compounds, etc.

PROBLEM PREVENTION

The Inlet: Setting A Maintenance Schedule

Many GC troubleshooting issues arise because system parts need to be replaced. It is not always obvious which part needs replacing. Therefore, a fair amount of time can be spent locating the problem part before it can be fixed.

Instead of waiting for a problem to occur, it is best to be proactive about your GC maintenance. For instance, the majority of GC issues are inlet related. By replacing inlet parts such as liners and septa on a regular basis, problems will occur far less frequently. Since problems will be occurring less often, there will be fewer instrument downtimes, resulting in greater productivity.

Below is a list of inlet parts which should be replaced regularly to prevent instrument downtime. Depending on how dirty the samples are, some parts will need to be replaced more or less often. In such instances, adjust the length of time or number of injections that is appropriate for the samples.

	Item Replacement Frequency	
	Septa	100 Injections (depends on needle style)
	Inlet Liner	Sample & Matrix Dependent Common Replacement Frequencies • Dirty/Soil Samples: < 2 weeks • Water Extracts: ~ 4 weeks • Headspace Extracts: ~ 6 months
00	O-Ring	6 months (or with each liner change)
	Inlet Seal	Sample Dependent (no more than 6 months)

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For a much more in-depth discussion of setting your GC inlet maintenance schedule, please contact your GC Specialist at GCSpecialist@Phenomenex.com.

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PROBLEM PREVENTION

The Inlet: Maintenance & Cleaning



Warning! This procedure involves the use of compressed gas and therefore eye protection should be worn.

Note: It best to have clean replacement liners or inserts available for quick exchange.

Full maintenance cleaning procedure:

- 1. Turn off inlet heat and allow inlet to cool.
- 2. Remove septum.
- 3. Remove liner or insert.
- 4. Remove base seal if applicable.
- 5. Use dry air or nitrogen to blow out any loose particles.
- 6. Use swab and solvent to clean interior walls if required.
- 7. Replace septum, liner or insert, and base seal.
- 8. Vent lines may also require replacement or cleaning.
- 9. Reassemble inlet and purge with clean, dry gas to remove solvent.

Note: Light maintenance may not require changing of septum or base seal. Avoid touching any parts that go inside the inlet with fingers as fingerprints will cause contamination.



PROBLEM PREVENTION

The Detector: Maintenance & Cleaning



Warning! Wear eye protection when working with fused silica tubing or compressed gas.

Electron Capture Detectors (ECD)

Because of the use of radioactive nickel in this type of detector, it should not be disassembled by those without specialized training and an appropriate license. Cleaning is limited to baking it out at 350 °C from 3 hours to overnight. Verify there are no leaks and the carrier gas is clean and dry before baking.

Flame Ionization Detectors (FID)

The collector bore and the jet require occasional cleaning to remove deposits. The deposits, which usually consist of white silica from column bleed or black carbonaceous soot, cause noise and spikes.

Cleaning procedure:

- 1. Turn off detector and its heater.
- 2. Turn off gases to the detector.
- 3. Allow time for the detector to cool.
- 4. Open up the detector and use mechanical means (brush, wire, etc., and compressed gas) to remove contamination.
- 5. Wash out the collector with distilled water and organic solvents as required.
- 6. Dry in an oven at about 70 °C for more than half an hour.

Flame Photometric Detectors (FPD)

Cleaning procedure:

- 1. Set instrument temperatures to cool to safe temperatures.
- 2. Turn off gasses to the detector.
- 3. Turn off power to the gas chromatograph and unplug main power cord.
- 4. Remove detector covers, disconnect, and remove the detector.
- 5. Remove and inspect jet assembly. Remove any deposits mechanically, for example, by using a wire.
- 6. Inspect and clean, if necessary, the glow plug and the guartz windows.
- 7. Blow loose particles away with compressed gas.
- 8. Replace the jet if it is damaged or difficult to clean with a wire.

Nitrogen Phosphorous Detectors (NPD)

Caution: If the hydrogen gas used to fuel the NPD detector is left on after the detector is disconnected from the column, this gas can accumulate in the oven and create an explosion hazard.

The collector bore and the jet require occasional cleaning to remove deposits. The deposits, which usually consists of white silica from column bleed or black carbonaceous soot, cause noise and spikes.

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COLUMN CARE & USE GUIDELINES



COLUMN CARE & USE GUIDELINES

Column Installation

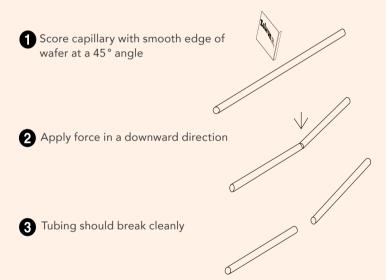
Pre-Installation Checklist

- Replace oxygen, moisture and hydrocarbon traps as necessary.
- Ensure that the injection port is clean and free of sample residues, septum, or capillary debris.
- Check and replace as necessary critical injector components such as seals, liners, and septa.
- Check and replace detector seals as necessary.
- Carefully inspect your column for damage or breakage.
- Check gas cylinder pressures to ensure that an adequate supply of carrier, make-up and fuel gases are available. Carrier gases should be of the highest purity. Note: It is critical that oxygen and water be removed from the carrier gas by the appropriate use of filters and adsorbents.

Designate A Flow Direction

Note: GC columns do not have a specific directional flow when received from the manufacturer. Upon initial use of your new Zebron™ column, Phenomenex recommends the practice of dedicating one specific end of the column for injector installation only. This is particularly important when dealing with active/caustic or contaminating compounds. If these compounds are routinely injected onto the column, degradation of the phase will occur leading to higher bleed. A typical first step to remedying (removing) this bleed would be to trim 10 cm from the front (injector) end of the column and keep trimming this inlet end of the column as necessary. Trying to remedy any bleed issues by trimming the column may not work if both ends have been interchangeably installed into the inlet.

Ensure A Proper Column Cut



Inspect cut with a magnifying glass - the cut should be smooth, not jagged



COLUMN CARE & USE GUIDELINES

Column Installation

Installation Into the Injector

- Place a capillary nut and ferrule on the injector end of the GC column, allowing a section of column to protrude.
 Trim one to two centimeters from the protruding end to remove ferrule contamination that may have entered the column. Inspect the cut with a magnifier to ensure that a smooth, clean, square-cut edge has been made recut if necessary.
- 2. Carefully hang the column in the GC oven, being cautious not to scratch or damage the polyimide coating on the capillary tubing. Rotate the column to avoid sharp bends of the capillary column and any contact of the column with oven surfaces.
- 3. Insert the column into the injector exactly the correct distance specified in the instrument manual. Tighten the ferrule nut finger-tight then 1/2 turn with a wrench. If the column can still be moved, tighten another 1/4 turn until the column is secure.
- 4. Adjust the carrier gas to obtain the flow rate listed on the test chromatogram.

Installation Into the Detector

Note: For users with sensitive detectors such as MS and ECD, column conditioning steps should be performed before installing the column to prevent contamination and frequent maintenance of the detector.

- 1. Place the column nut and ferrule past the end of the column and cut a centimeter or two off the end of the column. Be sure that the ferrule is the right size and pointing in the correct direction. Inspect the cut with a magnifier and ensure that the cut is square and smooth. Recut if needed.
- 2. Insert the outlet end of the column into the detector exactly the distance prescribed in the instrument manual. Distances will vary between detectors. Tighten the ferrule nut finger-tight then 1/2 turn with a wrench. If the column can still be moved, tighten another 1/4 turn until the column is secure.
- 3. Inspect the column connections for leaks using an electronic leak detector. Leaks at the inlet end may introduce oxygen to the column that will result in increased column bleed and damage to the column phase.

COLUMN CARE & USE GUIDELINES

Conditioning Basics: The Column

Column Conditioning Steps

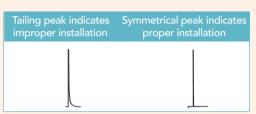
- 1. Allow sufficient time for the carrier gas to flow through the column to purge any oxygen that may be in the system.
- 2. Raise the temperature of the column to the maximum isothermal operating temperature that is listed on the individual Zebron™ GC Column Test Report. Maintain this temperature until a constant baseline is achieved. Conditioning times will depend on the phase identity and thickness, with thicker films taking longer to stabilize. In order to minimize the downtime of the instrument, columns can be conditioned overnight at the maximum isothermal temperature.

Installation Testing

- 1. Inject a detectable unretained sample, such as methane for an FID, to determine dead volume time and linear gas velocity at the desired column temperature. Adjust gas pressure for optimal flow depending on carrier gas selection.
- 2. The non-retained peak must have ideal peak shape or installation is faulty and needs to be redone.

If the peak is broad and/or tailing, check the following:

- Improper column positioning/insertion into inlet or detector
- Gross contamination of the splitter sleeve
- Chipped or cracked splitter sleeve
- Improper sweeping of sample at column end by makeup gas
- · Damaged or crushed column end



Unretained Peak Times and Markers

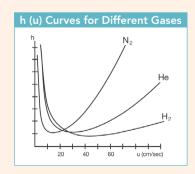
Methane with FID/TCD: Calculate linear velocity by injecting $25-100\,\mu\text{L}$ of 1% methane in N_2 gas blend. Measure the retention time of the methane peak and calculate the following:

Linear Velocity (u) = L/to

Detector Type	Marker Compound			
ECD	Methylene chloride ^{2, 3} , Dichlorodifluoromethane			
FID	Methane, Butane ¹			
NPD	Acetonitrile ^{2, 4}			
PID ELCDVinyl chloride				
TCD, MS Methane, Butane ¹ , air				
From a disposable lighter Place 1-2 drops in an autosampler vial and tightly cap. Shake and inject 1-2 μL from the headspace of the vial. Do not inject any liquid. Use a column temperature above 55 °C. Use a column temperature above 95 °C.				

Recommended Non-Retained Retention Times

Length (m)	H ₂ (sec)	He (sec)	N ₂ (sec)
15	38	75	150
30	75	150	300
60	150	300	600



COLUMN CARE & USE GUIDELINES

Conditioning Basics: The System

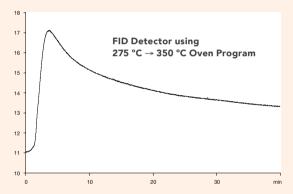
Increased signal is common and expected during the conditioning step when new components are installed. After a new column installation, detectors show increased signal response that decrease slowly over time with constant increased temperature. Misconception is that the baseline rise is solely due to column bleed. It may be a combination of many causes that collectively can be called system bleed. In fact, at temperatures below 200 °C, almost all background signal is the result of system noise.

Detector Effects

During new column installation, detectors are sometimes allowed to cool for convenience. If the column is connected to the detector during conditioning, the detector can become contaminated. When the detector is heated following column installation an increased signal will be observed that can be interpreted as column bleed as shown below. An increase in detector temperature from a constant temperature of 275 °C to 350 °C caused a signal increase of 6 pA!

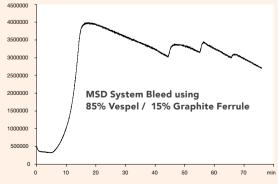
The effect would be greater if the detector had been constant at room temperature for an extended period of time. It is not uncommon for more sensitive detectors such as Electron Capture Detectors (ECD) to show very high signals that may persist for days after prolonged dormancy.

Repetitive heating and cooling of detectors can cause seal and ferrule distortion, allowing leaks to form. This might introduce oxygen into the system resulting in increased signal response.



Accessory Effects

If the detector temperature remains constant, other causes for baseline increases are still possible. Ferrules absorb gases and other substances that offgas when heated. Figure shows a spectrum that was obtained when an uncoated capillary column, which does not contain stationary phase, was installed using new ferrules. The signal is due to system noise only and not column bleed. Notice that the intensity of the signal is very high and would be greater than most analyte peaks. This would decrease the signal to noise ratio making detection limits much worse than if the system was conditioned and the baseline minimized.



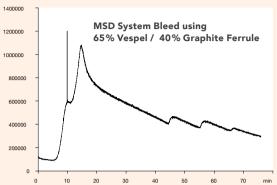
Oven Program:

40 °C for 5 min to 320 °C at 30 °C/min for 30 min to 340 °C at 30 °C/min for 10 min to 360 °C at 30 °C/min for 10 min to 370 °C at 30 °C/min for 10 min.

COLUMN CARE & USE GUIDELINES

Conditioning Basics: The System

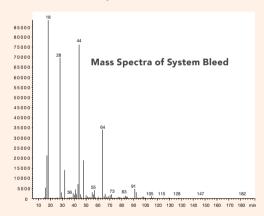
Ferrule composition also effects system bleed. After the completion of the same oven program using a ferrule with higher graphite content, the signal is 90% lower.



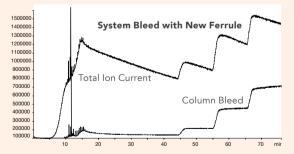
Oven Pregram:

40 °C for 5 min to 320 °C at 30 °C/min for 30 min to 340 °C at 30 °C/min for 10 min to 360 °C at 30 °C/min for 10 min to 370 °C at 30 °C/min for 10 min.

The major ions seen are 17, 18, 28, 32, and 44, and 64. Most masses can be easily explained by common gases adsorbing on the ferrule (such as water, carbon monoxide, nitrogen, oxygen, and carbon dioxide.) Since these same ions are indicators of a gas leak or contaminated vacuum chamber, an air and water check was run and passed before analysis. Subsequent runs after initial conditioning showed drastically reduced signals for these ions. An air leak would likely remain constant.



New 60/40% Vespel-Graphite ferrules were installed with a Zebron™ ZB-5 column. Subtracting the ions associated with system bleed shows bleed from column conditioning only. At normal operating conditions, over 90% of the ion intensity is completely due to system bleed derived from ferrule offgassing.



Installation of any column should be followed by a heating cycle to condition the system. During this conditioning cycle, the entire system is being conditioned, not only the column. To prevent detector contamination, it is recommended that you do not connect the column to the detector. Ferrule composition can also determine the extent of conditioning needed, as it is the key component responsible for most offgassing. Injector contributions should also be considered when determining system bleed, they may appear as peaks.



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Australia t: +61 (0)2-9428-6444 auinfo@phenomenex.com

Austria +43 (0)1-319-1301 anfrage@phenomenex.com

Belgium t: +32 (0)2 503 4015 (French) t: +32 (0)2 511 8666 (Dutch) beinfo@phenomenex.com

Canada t: +1 (800) 543-3681 info@phenomenex.com

t: +86 400-606-8099 cninfo@phenomenex.com

Czech Republic t: +420 272 017 077 cz-info@phenomenex.com

Denmark +45 4824 8048

nordicinfo@phenomenex.com

Finland

t: +358 (0)9 4789 0063 nordicinfo@phenomenex.com

France t: +33 (0)1 30 09 21 10 franceinfo@phenomenex.com

Germany t: +49 (0)6021-58830-0 anfrage@phenomenex.com

Hong Kong t: +852 6012 8162

hkinfo@phenomenex.com

India t: +91 (0)40-3012 2400 indiainfo@phenomenex.com

Indonesia

t: +62 21 5010 9707 indoinfo@phenomenex.com

t: +353 (0)1 247 5405 eireinfo@phenomenex.com

Italy t: +39 051 6327511 italiainfo@phenomenex.com

+81 (0) 120-149-262 jpinfo@phenomenex.com

Luxembourg t: +31 (0)30-2418700 nlinfo@phenomenex.com

Mexico t: 01-800-844-5226 tecnicomx@phenomenex.com

The Netherlands +31 (0)30-2418700

nlinfo@phenomenex.com

New Zealand t: +64 (0)9-4780951 nzinfo@phenomenex.com

Norway t: +47 810 02 005 nordicinfo@phenomenex.com

Poland +48 22 104 21 72 pl-info@phenomenex.com

Portugal t: +351 221 450 488

Singapore t: +65 800-852-3944 sginfo@phenomenex.com

Slovakia +420 272 017 077 sk-info@phenomenex.com

Spain t: +34 91-413-8613 espinfo@phenomenex.com

Sweden t: +46 (0)8 611 6950 nordicinfo@phenomenex.com

Switzerland

t: +41 (0)61 692 20 20 swissinfo@phenomenex.com

Taiwan t: +886 (0) 0801-49-1246 twinfo@phenomenex.com

Thailand t: +66 (0) 2 566 0287

thaiinfo@phenomenex.com

United Kingdom t: +44 (0)1625-501367 ukinfo@phenomenex.com

t: +1 (310) 212-0555 info@phenomenex.com

All other countries/regions Corporate Office USA t: +1 (310) 212-0555 info@phenomenex.com



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